




1 cellular_raza: Cellular Agent-based Modeling from a 2 Clean Slate

3 Jonas Pleyer ¹ and Christian Fleck¹

4 ¹ Freiburg Center for Data-Analysis and Modeling

DOI: [10.xxxxxx/draft](https://doi.org/10.xxxxxx/draft)

Software

- [Review](#) 
- [Repository](#) 
- [Archive](#) 

Editor: [Open Journals](#) 

Reviewers:

- [@openjournals](#)

Submitted: 01 January 1970

Published: unpublished

License

Authors of papers retain copyright
and release the work under a
Creative Commons Attribution 4.0
International License ([CC BY 4.0](#)).

5 Summary

6 cellular_raza is a cellular agent-based modeling framework which allows researchers to
7 construct models from a clean slate. In contrast to other agent-based modeling toolkits,
8 cellular_raza was designed to be free of assumptions about the underlying cellular repre-
9 sentation. This enables researchers to build up complex models while retaining full control
10 over every parameter introduced. It comes with predefined building blocks for agents and their
11 physical domain to quickly construct new simulations bottom-up. Furthermore, cellular_raza
12 can be used with the pyo3 and maturin packages and thus act as a numerical backend to a
13 python package.

14 Statement of Need

15 Agent-based models have become popular in cellular biology ([Cess & Finley, 2022](#); [Delile et al., 2017a, 2017b](#); [Mogilner & Manhart, 2016](#)). While these tools have proven to be effective
16 for targeted research questions, they often lack the ability to be applied for multiple distinct
17 use-cases in a more generic context. Nevertheless, core functionalities such as numerical
18 solvers, storage solutions, domain decomposition methods and functions to construct these
19 simulations could be shared between models if written generically. In order to address this issue
20 and construct models from first principles without any assumptions regarding the underlying
21 complexity or abstraction level, we developed cellular_raza.
22

23 State of Field

24 General-Purpose Agent-Based Modeling Toolkits

25 General-purpose agent-based toolkits are often designed without specific applications in mind
26 ([Abar et al., 2017](#); [Datseris et al., 2022](#); [Wilensky, 1999](#)). They are often able to define
27 agents bottom-up and can be a good choice if they allow for the desired cellular representation.
28 However, they lack the explicit forethought to be applied in cellular systems. Since they are
29 required to solve a wider range of problems they are not able to make assumptions on the
30 type of agent or the nature of their interactions and thus miss out on possible performance
31 optimizations and advanced numerical solvers.

32 Cellular Agent-Based Frameworks

33 In our previous efforts ([Pleyer & Fleck, 2023](#)), we assessed the overall state of modelling toolkits
34 for individual-based cellular simulations. The frameworks reviewed are all designed for specific
35 use cases and often require a large number of parameters which are often unknown in practice
36 and difficult to determine experimentally. This is an inherent problem for the applicability
37 of the software and the ability to properly interpret results. Few modelling frameworks exist

38 that provide a significant degree of flexibility and customisation in the definition of cell agents.
39 Chaste ([Cooper et al., 2020](#)) allows reuse of individual components of its simulation code, such
40 as ODE and PDE solvers, but is only partially cell-based. Biocellion ([Kang et al., 2014](#)) has
41 support for different cell shapes such as spheres and cylinders, but admits that their current
42 approach lacks flexibility in the subcellular description. BioDynaMo ([Breitwieser et al., 2021](#))
43 offers some modularity in the choice of components for cellular agents, but cannot freely
44 customise the cellular representation.

45 **cellular_raza**

46 We distinguish between different simulation aspects, e.g., mechanics, cell cycle, or cell cycle.
47 These aspects are directly related to the properties of the cells, domain, or other external
48 interactions. The user selects a cellular representation, which can be built from pre-existing
49 building blocks or a fully customised bottom-up approach, if desired. 'cellular_raza' utilises
50 macros to generate code contingent on the simulation aspects being solved numerically. It
51 makes extensive use of generics and provides abstract numerical solvers. 'cellular_raza' hides
52 the inherent complexity of the code generation process, yet enables users to modify the
53 specifics of the simulation through the use of additional keyword arguments within the macros.
54 Consequently, users are able to fully and deeply customise the representation and behaviour
55 of the agents. Each simulation aspect is formulated as a trait in Rust's type system, which
56 provides the necessary abstractions. The [getting-started](#) guide provides a good entry point
57 and explains every step from building, running to visualising.

58 **Examples**

59 In the following, we present four different examples how to use cellular_raza (see [cellular-
60 raza.com/showcase](#)).

61 **Cell Sorting**

62 Cell sorting is a naturally occurring phenomenon ([Graner & Glazier, 1992](#); [Steinberg, 1963](#)).
63 While the underlying biological reality can be quite complex, it is rather simple to describe
64 such a system in its most basic form. Fundamentally, any cellular Interaction is specific to
65 their species. We consider two distinct species represented by soft spheres which physically
66 attract each other at close proximity if their species is identical. Cells are placed randomly
67 inside a cube with reflective boundary conditions. In the final snapshot, we can clearly see the
68 phase-separation between the different species.

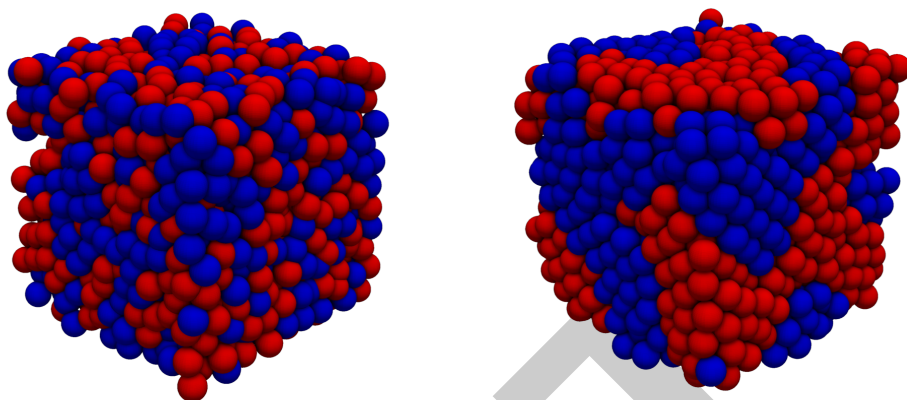


Figure 1: The initial random placement of cells reorders into a phase-separated spatial pattern.

69 Bacterial Rods

70 Bacteria come in various forms (Young, 2006; Zapun et al., 2008) such as elongated shapes
71 (Billaudeau et al., 2017) which grows asymmetrically in the direction of elongation. Our model
72 describes the physical mechanics of one cell as a collection of multiple vertices \vec{v}_i which are
73 connected by springs. Their relative angle α at each connecting vertex introduces a stiffening
74 force which is proportional to $2 \tan(\alpha/2)$. Cells interact via a soft-sphere force potential with
75 short-ranged attraction. Multiple contributions are calculated between every vertex and the
76 closest point on the other cells edges. In addition, the cell cycle introduces growth of the
77 bacteria until it divides in the middle into two new cells. This growth is downregulated by an
78 increasing number of neighboring cells which is a phenomenological but effective choice for
79 the transition into the stationary phase of the bacterial colony. Cells are placed inside the
80 left-hand side of an elongated box with reflective boundary conditions. Their colors range from
81 green for fast growth to blue for dormant cells.

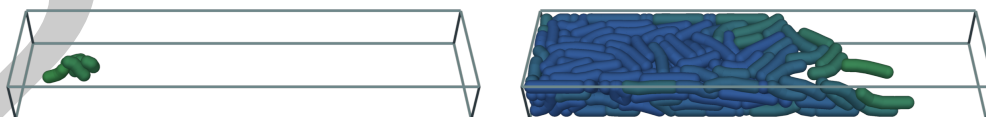


Figure 2: The bacteria extend from the initial placement in the left side towards the right side. Their elongated shape and the confined space favour the orientation facing along the growth direction.

82 **Branching of *Bacillus Subtilis***

83 Spatio-temporal patterns of bacterial growth such as in *Bacillus Subtilis* have been studied for
84 numerous years (Kawasaki et al., 1997; Matsushita et al., 1998). Cells are modeled by soft
85 spheres which interact with the domain by taking up nutrients. By consuming intracellular
86 nutrients, the cell grows continuously and divides upon reaching a threshold. The initial
87 placement of the cells is inside of a centered square. From there, cells start consuming nutrients
88 and growing outwards towards the nutrient-rich area. Cells are colored bright purple while they
89 are actively growing and dividing while dark cells are not subject to growth anymore. The
90 outer domain is colored by the intensity of present nutrients. A lighter color indicates that
91 more nutrients are available while a dark color signifies a lack thereof.

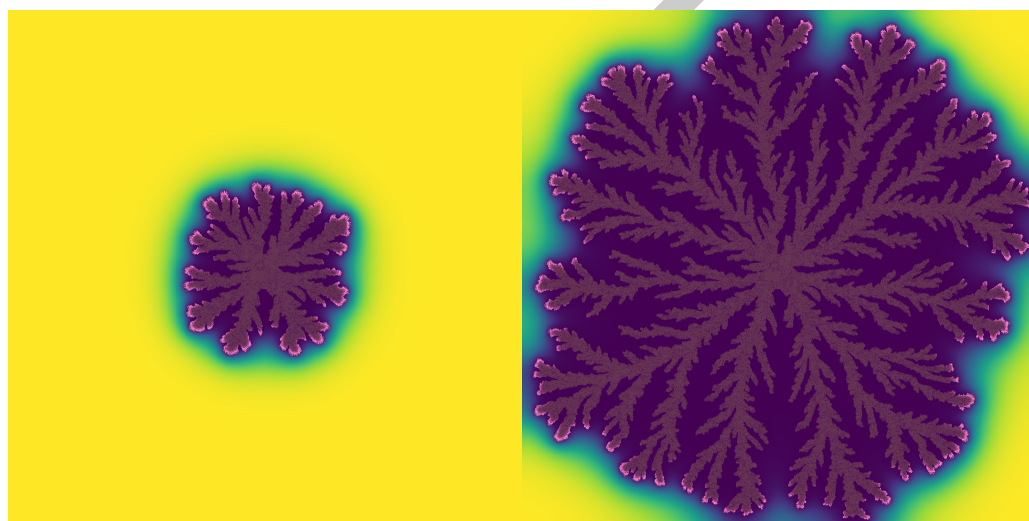


Figure 3: The bacterial colony grows outwards towards the nutrient-rich parts of the domain thus forming branches in the process.

92 **Semi-Vertex Model for Epithelial and Plant Cells**

93 Vertex models are a very popular choice in describing multicellular systems. They are actively
94 being used in great variety such as to describe mechanical properties of plant cells (Merks et
95 al., 2011) or organoid structures of epithelial cells (Barton et al., 2017; Fletcher et al., 2014).

96 We represent cells by a polygonal collection of vertices connected by springs. An inside pressure
97 pushes vertices in an outwards direction. These two mechanisms by themselves create perfect
98 hexagonal cells. Cells are attracting each other but in the case where two polygons overlap,
99 a repulsive force acts between them. Cells are placed in a perfect hexagonal grid such that
100 edges and vertices align. Their growth rates are chosen from a uniform distribution.

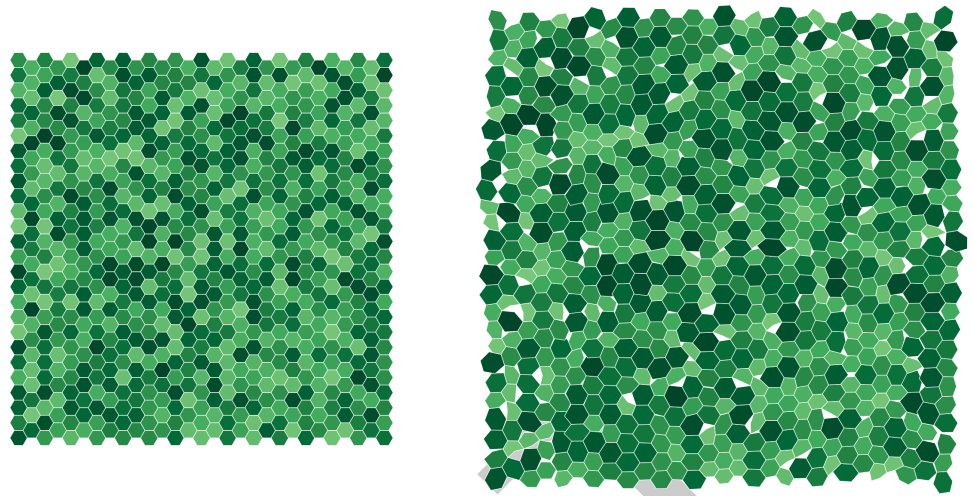


Figure 4: During growth the cells push on each other thus creating small spaces in between them as the collection expands. These forces also lead to deviations in the otherwise perfect hexagonal shape.

Acknowledgements

101

102 The author(s) declare that financial support was received for the research, authorship, and/or
103 publication of this article. JP and CF received funding from FET-Open research and innovation
104 actions grant under the European Union's Horizon 2020 (CyGenTiG; grant agreement 801041).

References

105

- 106 Abar, S., Theodoropoulos, G. K., Lemariner, P., & O'Hare, G. M. P. (2017). Agent based
107 modelling and simulation tools: A review of the state-of-art software. *Computer Science*
108 *Review*, 24, 13–33. <https://doi.org/10.1016/j.cosrev.2017.03.001>
- 109 Barton, D. L., Henkes, S., Weijer, C. J., & Sknepnek, R. (2017). Active vertex model for
110 cell-resolution description of epithelial tissue mechanics. *PLOS Computational Biology*,
111 13(6), e1005569. <https://doi.org/10.1371/journal.pcbi.1005569>
- 112 Billaudeau, C., Chastanet, A., Yao, Z., Cornilleau, C., Mirouze, N., Fromion, V., & Carballido-
113 López, R. (2017). Contrasting mechanisms of growth in two model rod-shaped bacteria.
114 *Nature Communications*, 8(1). <https://doi.org/10.1038/ncomms15370>
- 115 Breitwieser, L., Hesam, A., Montigny, J. de, Vavourakis, V., Iosif, A., Jennings, J., Kaiser,
116 M., Manca, M., Di Meglio, A., Al-Ars, Z., Rademakers, F., Mutlu, O., & Bauer, R.
117 (2021). BioDynaMo: a modular platform for high-performance agent-based simulation.
118 *Bioinformatics*, 38(2), 453–460. <https://doi.org/10.1093/bioinformatics/btab649>
- 119 Cess, C. G., & Finley, S. D. (2022). Multiscale modeling of tumor adaption and invasion
120 following anti-angiogenic therapy. *Computational and Systems Oncology*, 2(1). <https://doi.org/10.1002/cso2.1032>
- 122 Cooper, F., Baker, R., Bernabeu, M., Bordas, R., Bowler, L., Bueno-Orovio, A., Byrne, H.,
123 Carapella, V., Cardone-Noott, L., Cooper, J., Dutta, S., Evans, B., Fletcher, A., Grogan, J.,
124 Guo, W., Harvey, D., Hendrix, M., Kay, D., Kursawe, J., ... Gavaghan, D. (2020). Chaste:
125 Cancer, heart and soft tissue environment. *Journal of Open Source Software*, 5(47), 1848.
126 <https://doi.org/10.21105/joss.01848>

- 127 Datseris, G., Vahdati, A. R., & DuBois, T. C. (2022). Agents.jl: A performant and
128 feature-full agent-based modeling software of minimal code complexity. *SIMULATION*,
129 003754972110688. <https://doi.org/10.1177/00375497211068820>
- 130 Delile, J., Herrmann, M., Peyri ras, N., & Doursat, R. (2017a). A cell-based computational
131 model of early embryogenesis coupling mechanical behaviour and gene regulation. *Nature*
132 *Communications*, 8(1). <https://doi.org/10.1038/ncomms13929>
- 133 Delile, J., Herrmann, M., Peyri ras, N., & Doursat, R. (2017b). A cell-based computational
134 model of early embryogenesis coupling mechanical behaviour and gene regulation. In
135 *Nature Communications* (No. 1; Vol. 8). Springer Science; Business Media LLC. <https://doi.org/10.1038/ncomms13929>
- 136
- 137 Fletcher, A. G., Osterfield, M., Baker, R. E., & Shvartsman, S. Y. (2014). Vertex models of
138 epithelial morphogenesis. *Biophysical Journal*, 106(11), 2291–2304. <https://doi.org/10.1016/j.bpj.2013.11.4498>
- 139
- 140 Graner, F., & Glazier, J. A. (1992). Simulation of biological cell sorting using a two-dimensional
141 extended potts model. *Physical Review Letters*, 69(13), 2013–2016. <https://doi.org/10.1103/physrevlett.69.2013>
- 142
- 143 Kang, S., Kahan, S., McDermott, J., Flann, N., & Shmulevich, I. (2014). Biocellion :
144 Accelerating computer simulation of multicellular biological system models. *Bioinformatics*,
145 30(21), 3101–3108. <https://doi.org/10.1093/bioinformatics/btu498>
- 146 Kawasaki, K., Mochizuki, A., Matsushita, M., Umeda, T., & Shigesada, N. (1997). *Modeling*
147 *Spatio-Temporal Patterns Generated by Bacillus subtilis*. <https://doi.org/10.1006/jtbi.1997.0462>
- 148
- 149 Matsushita, M., Wakita, J., Itoh, H., R fols, I., Matsuyama, T., Sakaguchi, H., & Mimura, M.
150 (1998). *Interface growth and pattern formation in bacterial colonies*. [https://doi.org/10.1016/S0378-4371\(97\)00511-6](https://doi.org/10.1016/S0378-4371(97)00511-6)
- 151
- 152 Merks, R. M. H., Guravage, M., Inz , D., & Beemster, G. T. S. (2011). VirtualLeaf: An
153 open-source framework for cell-based modeling of plant tissue growth and development.
154 *Plant Physiology*, 155(2), 656–666. <https://doi.org/10.1104/pp.110.167619>
- 155 Mogilner, A., & Manhart, A. (2016). Agent-based modeling: Case study in cleavage furrow
156 models. *Molecular Biology of the Cell*, 27(22), 3379–3384. <https://doi.org/10.1091/mbc.e16-01-0013>
- 157
- 158 Pleyer, J., & Fleck, C. (2023). Agent-based models in cellular systems. *Frontiers in Physics*,
159 10. <https://doi.org/10.3389/fphy.2022.968409>
- 160 Steinberg, M. S. (1963). Reconstruction of tissues by dissociated cells: Some morphogenetic
161 tissue movements and the sorting out of embryonic cells may have a common explanation.
162 *Science*, 141(3579), 401–408. <https://doi.org/10.1126/science.141.3579.401>
- 163 Wilensky, U. (1999). *NetLogo* [Http://ccl.northwestern.edu/netlogo/]. Center for Connected
164 Learning; Computer-Based Modeling. <http://ccl.northwestern.edu/netlogo/>
- 165 Young, K. D. (2006). The selective value of bacterial shape. *Microbiology and Molecular*
166 *Biology Reviews*, 70(3), 660–703. <https://doi.org/10.1128/mnbr.00001-06>
- 167 Zapun, A., Vernet, T., & Pinho, M. G. (2008). The different shapes of cocci. *FEMS*
168 *Microbiology Reviews*, 32(2), 345–360. <https://doi.org/10.1111/j.1574-6976.2007.00098.x>